

Guidelines for the Genotyping of Mice and Rats

Purpose

The proper identification of genetically engineered animals in a litter is critical to the efficient pursuit of research and in reducing the number of animals involved in a research project. Most often the genotype is determined by analysis of DNA extracted from tissues of young rodents. Analysis by the Polymerase Chain Reaction (PCR) requires the least amount of DNA. DNA for PCR analysis can be obtained from ear punches, hair or fecal samples, oral or rectal swabs (1-9). Depending on the requirements of the study, investigators are urged to consider these noninvasive alternatives. Larger amounts of DNA are required for Southern Blot determination of the genotype. Obtaining tissue from a mouse or rat for DNA analysis via tail biopsy is a safe, effective and humane procedure. When performed properly it causes only minimal or transient pain and distress, and induces no more “physiological impact” (change in heart rate, body temperature, or activity level) than just restraining the animal for the procedure. (11) DNA prepared from tail biopsies is suitable for analysis by either Southern Blot or PCR.

Guidelines for Tail Biopsy

1. Procedures for tail biopsy for DNA analysis and/or genotyping must be described in an approved Animal Study Proposal (ASP).
2. Ideally, mice and rats should be **10-21** days old. At this age, the tail tissue is soft (vertebra are not yet calcified) and the yield of DNA is highest (8,10). In addition, prompt analysis of tail tissue allows the desired mice and rats to be identified prior to weaning which can facilitate more efficient use of cage space.
 - a. **For mice and rats 10-21 days of age:** Because pain sensory development may be complete, and to further minimize any transient pain or distress, investigators are strongly encouraged to apply local anesthesia to the tail. Local anesthesia may be achieved by immersion of the tail in ice cold ethanol for 10 seconds, by an application of ethyl chloride spray or by the use of another suitable anesthetic as recommended by the attending veterinarian.
 - b. **For mice and rats greater than 21 days of age:** The use of a local or general anesthetic is required prior to collection of tissue. If a general anesthetic is to be used, an appropriate agent should be recommended by the attending veterinarian.
 - c. **For rats greater than 35 days of age:** The use of a general anesthetic is required.
3. Manually restrain the mouse or rat between thumb and forefinger. This is a convenient time to identify the animals using the appropriate method (i.e. ear punch, ear tag, transponder etc.).
4. With sterile scalpel, razor blade, or scissors cleanly excise the distal 2mm (maximum 5 mm) of the tail. If the proper procedures are followed, the yield of DNA from 5 mm of tail should exceed 50 micrograms, enough for multiple analyses. The yield of DNA does not proportionally increase as tail fragments larger than 5mm are used. If

small amounts of DNA are required, investigators should consider taking only 2 mm of tail. If the analysis of the DNA is to be performed by PCR, great care should be taken to remove all tissue from the scissors or scalpel after each animal. Disinfect the scalpel or scissors between animals. If a scalpel is used, also disinfect the work surface on which the tail is placed between animals.

5. The investigator must monitor the animals to assure hemostasis after the animals are returned to the cage. If needed, apply digital pressure, silver nitrate, or other means of hemostasis.
6. If additional DNA is needed for retesting alternatives to a second tail biopsy should be considered (11). Repeat tail biopsies require anesthesia and must be justified in the ASP. The use of post-procedural analgesia should be considered.

References

1. Hofstetter JR, Zhang A, Mayeda AR, Guscar, T, Nurnberger JI and Lahiri DK. Genomic DNA from Mice: A Comparison of Recovery Methods and Tissue Sources. *Biochem Mol Med* 1997 Dec; 62(2):197-202.
2. Dennis, MB. IACUC Review of Genetic Engineering. *Lab Animal* 2000 Mar; 29(3):34-37
3. Irwin MH, Moffatt RJ and Pinkert CA. Identification of Transgenic Mice by PCR Analysis of Saliva. *Nat Biotechnol* 1996 Sep;14(9): 1146-8.
4. Schmitteckert EM, Prokop CM and Hedrich HJ. DNA Detection in Hair of Transgenic Mice - A Simple Technique Minimizing the Distress on the Animals. *Laboratory Animals* 1999; 33/4: 385-389.
5. Couse JF, Davis VL, Tally WC and Korach KS. An Improved Method of Genomic DNA Extraction for Screening Transgenic Mice. *National Institute of Environmental Health Sciences, National Institutes of Health. BioTechniques* 1994; 17:1030-1032.
6. Malumbres M, Mangues R, Ferrer N, Lu S and Pellicer A. Isolation of High Molecular Weight DNA for Reliable Genotyping of Transgenic Mice. *BioTechniques* 1997; 22/6:1114-1119.
7. Broome RL, Feng L, Zhou Q, Smith A, Hahn N, Matsui SM, Omary MB. Non-invasive Transgenic Mouse Genotyping Using Stool Analysis. *FEBS Lett* 1999; 462:159-160.
8. Pinkert CA. Transgenic Animal Technology: Alternatives in Genotyping and Phenotyping. *Comp Med* 2003; 53/2:126-139.
9. Meldgaard M, Bollen PJA, Finsen B. Non-invasive method for sampling and extraction of mouse DNA for PCR. *Laboratory Animals* 2004; 38:413-417.
10. Shinohara H. The Musculature of the Mouse Tail is Characterized by Metameric Arrangements of Bicipital Muscles. *Okajimas Folia Anat Jpn* 1999; 76:157-169
11. Cinelli P., et.al. Comparative Analysis and Physiological Impact of Different Tissue Biopsy Methodologies Used for the Genotyping of Laboratory Mice. *Lab Animals* 2007; 41: 174-184

Approved - 6/12/02

Revised - 1/12/05

Revised- 9/12/07